

Hanne C. Winther-Larsen
CURRICULUM VITAE

NAME: Hanne Cecilie Winther-Larsen
DATE OF BIRTH: 26/05/66
SEX: Female
NATIONALITY: Norwegian

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EDUCATION:

| | | |
|----------|---------------------------------------------------------------------------|-----------------------------------------|
| 2014 | : University of Oslo (UiO), Oslo, Norway | Science communication |
| 2010 | : University of Oslo, Oslo, Norway | Universitetspedagogisk basiskompetanse. |
| 2009 | : Umeå University, Umeå, Sweden | Microscopy in Microbial Systems. |
| 2007 | : University of Bergen, Bergen, Norway | From Innate to Specific Immunity. |
| 2001 | : University of Oslo, Oslo, Norway | Transmission Electron Microscopy. |
| 1993-99: | Norwegian University of Science and Technology (NUST), Trondheim, Norway. | Ph.D. in Biotechnology. |
| 1992-93: | University of Leicester, Leicester, England. | M.Sc. in Molecular Genetics. |
| 1988-92: | Heriot-Watt University, Edinburgh, Scotland. | B.Sc. (hon) in Microbiology. |
| 1988 | : University of Trondheim, Trondheim, Norway. | Cell biology |
| 1987 | : University of Oslo, Oslo, Norway. | Computer Science and Chemistry. |
| 1986 | : San Diego State University, California, USA. | English and Art. |
| 1985 | : University of Oslo, Oslo, Norway. | Philosophy. |

WORK EXPERIENCE:

2017-present :Head of Section of Pharmacology and Pharmaceutical Biosciences, Department of Pharmacy, UiO. Responsible for 38 employees. Reporting to Head of Department.

2011-2017 : Deputy Head of Department of Pharmaceutical Biosciences, UiO, Oslo, Norway.

2016 - : Professor in molecular microbiology, Department of Pharmacy, UiO, Oslo, Norway

2010 - 2016 : Associate Professor, School of Pharmacy, UiO, Oslo, Norway.

2009-2010 : Senior Research Scientist, Norwegian School of Veterinary Sciences (NVH), Oslo.

Fall 2008 : Visiting Scientist, Harvard Medical School, Boston, MA, USA.

2004 -2008 : Senior Scientist, Department of Molecular Biosciences (IMBV), UiO, Oslo.

Fall 2002 : Post-doc, Institute of Bacteriology, University of Utrecht, Utrecht, The Netherlands.

1999-03 : Post-doc, Biotechnology Centre of Oslo, UiO, Oslo.

1993-99 : Ph.D. studentship at UNIGEN Centre for Molecular biology, NUST, Trondheim.

1994-97 : Research assistant in molecular biology and biochemistry, NUST, Trondheim.

1984 : Laboratory Assistant at the Norwegian Research Centre for National Health, Oslo.

Prizes/ Awards/ Academy memberships:

2015-present: International Society of Extracellular vesicles.

2012-present: Norwegian Microbiology Society.

2002-present: Member American Society of Microbiology.

1996-present: Norwegian Biochemical Society,

2008: Publication prize at IMBV, UiO.

2007: Cover picture of J Bacteriol Issue 18, 189.

2006: Publication prize at IMBV, UiO.

2005: Publication prize at IMBV, UiO.

2001: Prize for best publication at School of Pharmacy, UiO.

GRANTS:

2020: Research Council of Norway (RCN) Milepæl grant. Project leader. Outer membrane vesicle vaccine against enteric redmouth disease (ERM) in aquaculture. Project 313638.

2019: Research Council of Norway (RCN) Havbruk2 program. Partner in project: Rational design of vaccination strategies for Atlantic cod (VACSACOD). Project 301965.

2019: Research Council of Norway INPART program. MISC. Project partner. Norwegian-Chinese-Japanese student and researcher exchange program on study of microbial surfaces.

2017: RCN Havbruk2 program: Project leader. Bacterial membrane vesicles: toxic or an improvement to fish health. Project 281056

2017: ERA-net MareLife: Work-package leader. The project PROMiSE.

2016: RCN FORNY program: Deputy head. 267845 Novel vaccine for salmon farming with UiO TTO Inven2 as project leader.

2016: RCN Havbruk2 program: Project leader. 268021 Implication of bacterial membrane vesicles for fish health in aquaculture.

2014: RCN FriMedBio program. Deputy head of project. Nanowire-mediated electron iron acquisition in Bacteria. Project 240741/CO.

2013: RCN Biotek2021 program. Project leader. Vaccination against intracellular bacterial pathogens for aquaculture. Project 233849.

2012: DAADppp program through RCN. Project leader. Dissection of Francisella virulence in a genetically tractable host system.

2012: Research Council of Norway. Havbruk program. Deputy head of project: Coordinated bacterial virulence: relevance in winter ulcer.

2009: Norwegian School of Veterinary Sciences. Internal 4 year PhD fellowship for Espen Brudal to explore Type IV pili in fish pathogenic *Francisella* species.

2009: Trippel alliance research grant in collaboration with the Veterinary Institute and University of Life Sciences studying transformation capabilities of fish pathogenic *Francisella* species.

2008: EMBIO travel grant in support of visiting Harvard Medical School, studying bacteriophage interaction with Type IV pili.

2008: Kristine Bonneveis travel grant for female scientist. Visiting Harvard Medical School, studying bacteriophage interaction with Type IV pili.

2002: Short-term EMBO fellowship (ASTF 139.00-02), visiting the University of Utrecht, in The Netherlands. Studying microbial adherence to human cells, and learning immunofluorescence- and immunogold electron-microscopy techniques.

2002: The Norwegian Cancer Society, Grete Harbitz og Lillemer Grobstok's stipend, in support of my scientific work.

2001: Henrik Homans Minde, in support of my scientific work.

1999 – 2002: Norwegian Research Council (NFR) post-doc stipend (128374/310). A novel pathway for Type IV pilus biogenesis with implications for organelle structure and function.

MOBILITY (research stays abroad lasting more than three months)

2008 Visiting scientist, Harvard Medical School, Boston, MA, USA.

2002 Visiting scientist, University of Utrecht, in The Netherlands

TEACHING :

FARM2140 Pharmaceutical microbiology, Department of Pharmacy, UiO. Since 2019.

FRM3010 Lecturing in Microbial quality control. Department of Pharmacy, UiO. Since 2010.

FRM9905 Course coordinator in a newly established course within the National PhD Research School in Pharmacy (NFIF). Since 2014. Molecular Microbiology in Pathogenesis and Evolution.

FRM1040 Course coordinator in Basic Biology, School of Pharmacy, UiO. Since 2012.

FRM2020 and FRM3010 Lecturing in Pharmaceutical Microbiology, School of Pharmacy. UiO. Since 2010.

MBV1010 Lecturing in Prokaryote Molecular Biology, Department of Molecular Biosciences, UiO. 2011.

MBV3060 Lecturing in Gene technology, Department of Molecular Biosciences, UiO. 2009.

FRM2010 Lecturing in Recombinant DNA technology, School of Pharmacy, UiO. Since 2002.

Laboratory course instructor in Molecular Biology and Biochemistry, Institute of Biotechnology, NUST. 1994-1997.

SUPERVISION:

Supervision of project students, summer students, diploma students, Erasmus students, Ph.D. students and research fellows.

Post-docs:

| | | | | |
|-----------|--------------------|------------|-----------------|-----------------|
| 2020 | Jan Janouskovec | post-doc | main supervisor | FAI, ERA-net |
| 2019-2020 | Elia Ciani | researcher | main supervisor | FAI, RCN |
| 2019 | Agnieszka Wrobel | researcher | main supervisor | FAI, RCN |
| 2018-2021 | Katrine Lekang | post-doc | main supervisor | FAI |
| 2017-2018 | Julia Tandberg | researcher | main supervisor | FAI, RCN |
| 2015-2017 | Leidy Lagos | post-doc | main supervisor | FAI, RCN |
| 2014-2016 | Alexander Kashulin | researcher | main supervisor | NMBU, RCN |
| 2012-2014 | Christian Karlsen | post-doc | co-supervisor | NMBU, RCN |
| 2009 | Christin Vanberg | post-doc | main supervisor | NSVS |
| 1999 | Sangeeta Bharati | post-doc | co-supervisor | UNIGEN, Pharmaq |

PhD students:

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|-----------|--------------------------------|-------|------------------|----------|
| 2020- | Gabriela Paz Carril Leiva, PhD | | co-supervisor | NMBU |
| 2019- | Verena Mertens | PhD | main supervisor | FAI, UiO |
| 2015-2019 | Anne Vogel | PhD | co-supervisor | NTNU |
| 2015-2020 | Petter Langlete | PhD | main supervisor | MN, UiO |
| 2013-2017 | Julia I. Tandberg | Ph.D. | main supervisors | FAI, UiO |
| 2011-2015 | Marianne Arnemo | Ph.D. | co-supervisor | MLS, UiO |
| 2011-2015 | Arturas Kavaliauskis | Ph.D. | co-supervisor | UiO |
| 2010-2015 | Elisabeth O. Lampe | Ph.D. | main supervisor | FAI, UiO |
| 2009-2014 | Espen Brudal | Ph.D. | main supervisor | NMBU |
| 2003-2006 | Cecilia Løvold | Ph.D. | co-supervisor | UiO |

MSc students:

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|------------|----------------------|-------|-----------------|---------------------------|
| 2019-2020 | Ahmed Ek | M.Sc. | main supervisor | FAI, UiO |
| 2019-2020 | Rebekka Rolfsnes | M.Sc. | main supervisor | IBV, UiO |
| 2018-2019 | Chirine Issam Kanaan | M.Sc. | main supervisor | FAI, UiO |
| 2017-2018 | Sanja Bogdanovic | M.Sc. | main supervisor | FAI, UiO |
| 2017-2018 | Mwisa Amani Bulere | M.Sc. | main supervisor | FAI, UiO |
| 2014-2015 | Diana Truong | M.Sc. | main supervisor | FAI, UiO |
| 2013-2014 | Eva Berger | M.Sc. | main supervisor | IBV, UiO |
| 2013-2015 | Tone Lian | M.Sc. | co-supervisor | IBV, UiO |
| 2012-2013 | Dai Uy Truong | M.Sc. | main supervisor | FAI, UiO |
| 2012-2013 | Ida Marie Thrane | M.Sc. | main supervisor | FAI, UiO |
| 2011- 2012 | Hanh Thi Hieu Phan | M.Sc. | main supervisor | FAI, UiO |
| 2010- 2011 | Anne Paulenz Hope | M.Sc. | co-supervisor | IMBV, UiO |
| 2009-2010 | Kristian Franer | M.Sc. | main supervisor | NSVS |
| 2007-2008 | Raimonda Butkute | M.Sc. | co-supervisor | IMBV, UiO |
| 2001-2003 | Cecilia Løvold | M.Sc. | co-supervisor | Biotechnology Center, UiO |
| 1996 | Bjørnar Valand | M.Sc. | co-supervisor | NTNU |

International students, Summer student Erasmus+ students and visting scientists:

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|------------|--------------------|-----------------|---------------|-------------------------------|
| 2017, 2019 | Cristian Oliver | Post-doc | supervisor | Universidad de Austral, Chile |
| 2018 | Nangialai Ahmadzai | Summer student, | supervisor | UiO |
| 2017 | Sara Ahmed | Erasmus+ | supervisor | Uni of Portsmouth, UK |
| 2015 | Sheemal Kumar | PhD | co-supervisor | MacQuarie Uni, Sydney |

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|------------|-------------------|-----|---------------|------------------------|
| 2013 -2014 | Yannick Brenz | PhD | co-supervisor | BNI, Hamburg, Germany. |
| 2005 | Emilie Salmonsson | PhD | co-superviors | Umeå Uni, Umeå. |

ORGANIZATIONAL EXPERIENCE:

2020 : Co-organization of the Strategic leader team seminar of MN Faculty, UiO.
 2013-2018 : Leader team of the MN faculty, UiO, Top-Tier Initiative, Centre of Intergrative Microbial Evolution (CIME). <http://www.mn.uio.no/ibv/english/research/groups/cime/>
 2012-2014 : Leader of The Norwegian Microbiology Society. <http://www.nfmikro.net/>
 2005 - 2006 : Assistant platform leader, The FUGE platform CAMST: Consortium of Advances Microbial Sciences and Technologies. (<http://camst.org>)
 Fall 2001 : Chairman of the Biochemical Interest Group, Oslo.
 Spring 2001 : Vice-chairman of the Biochemical Intererst Group, Oslo.

Scientific meetings:

2015 : Co-organizer and scientific committee member of 9th European Zebrafish Meeting (9EZM) Oslo, Norway, June 28-July 2, 2015. <http://www.ccnorway.no/zebrafish/>.
 2014-present : Organizer of the CIME bi-weekly seminar series.
 2011 : Co-organizer of ETOX 15, a European Workshop on Microbial Protein Toxins.
 2010-2011 : Organizer of the MicroNET seminar series (Consortium of microbiologist at Mathematical and Natural Science Faculty, UiO)
 2006 : Organizing the 1st student/post-doc meeting in Microbiology in Norway (NoMi-06).

Board representative:

2014-2017 : Faculty member in educational board for MSc in Pharmacy, UiO.
 2013-2018 : Board member of Centre of Excellence, Centre for Ecological and Evolutionary Synthesis (CEES), UiO. <http://www.mn.uio.no/cees/english/>
 2005 -2006 : Board member at IMBV, UiO.
 2005 -2006 : Member of IMBV strategic committee for 2005 - 2009. UiO.
 2005 - 2006 : Member of IMBV's appointment committee. UiO
 2002 : Board member at The Biotechnology Center of Oslo. UiO.

COMMISSIONS OF TRUST

2019 : Reviewer for Netherland Research Council NWO ISAAC.
 2018 : Evaluator with a panel of six for the RCN Young Scientist of the Year Award for the Medical & Biological Sciences track.
 2017-present : Evaluators of Associate professor and professor positions at NUST and Norwegian University of Life Sciences.
 2013-present : Acted as PhD thesis opponent at Norwegian University of Life Sciences and NUST.
 2011, 2013 : Reviewer in French research council ANR
 2011 : External reviewer for Icelandic Research Fund. 2011.
 2011-present : Reviewer of research articles in Microbiology (The Society for General Microbiology), PLOS One, Journal of Applied Microbiology, Journal of Fish Diseases, Veterinary Microbiology, Scientific Reports

PATENTS/DOFI FILED:

2020 : Use of Outer Membrane Vesicles against yersiniosis.
 2015 : Enhanced growth media for *Piscirickettsia salmonis*
 2014 : Attenuated of the fish pathogenic *Francisella noatunensis* subsp *noatunensis* for vaccination against francisellosis.
 2013 : Patentability Analysis of Disclosure 13020: *Francisella noatunensis* Outer Membrane Vesicles and uses Thereof.
 2012 : DOFI filed - Fluorescent strains of fish pathogenic Francisella.
 1999 : Patent filing – British patent application no. 9910413.5- Pm MUTANTS

1999 : Patent filing – New british patent application – pH Control of Expression

MAJOR COLLABORATIONS

- Prof Anders Sjöstedt, Francisella genetics, Umeå University and Molecular Infections Medicine Sweden (MIMS), Umeå, Sweden
- Prof Ian Paulsen, Macquarie University, Sydney, Australia
- Senior lecturer Maria Liaskos, La Trobe University, Melbourne, Australia.
- Prof Alejandro Yanez, Universidad de Austral de Chile, Valdivia, Chile.
- Prof. Gareth Griffiths, Zebrafish as an infection model for bacterial infections in aquaculture. Department of Biosciences, UiO, Norway
- Prof. Henning Sørum, Fish pathogen and host interaction, Norwegian University of Life Sciences, Norway.
- Senior scientist Duncan Colquhoun, Norwegian Veterinary Institute, Norway
- Dr. Håvard Sletta, High-throughput-screening of microbial phenotyping, SINTEF, Trondheim, Norway
- Prof. Bente Edvardsen, Bioprospecting of the Nordic Algal Collection for Antimicrobial compounds, Department of Biosciences, UiO, Norway.
- Prof. Hanne Hjorth Tønnesen, Department of Pharmacy, UiO, Norway.
- Prof. Dirk Linke, Department of Biosciences, UiO, Norway
- Senior researcher Dr Duncan Colquhoun, Norwegian Veterinary Institute, Norway
- Prof. Martin Homann-Marriot, Bacterial nanowires, Norwegian University of Science and Technology, Trondheim, Norway.

SCIENTIFIC EVALUATOR:

Sensor and committee member of M.Sc (15) and Ph.D (10) thesis defences.

Invited presentations to peer-reviewed, internationally established conferences and/or international advanced schools;

- 2019 Speaker at International Society for Extracellular Vesicles ISEV19, Kyoto, Japan, 23-30 April 2019.
- 2019 Speaker at 3rd Fish and Shellfish Immunology Conference, Las Palmas, Gran Canaria, 16-20 June.
- 2018 Plenary speaker at OMV vaccine works-shop, Varadero, Cuba, 10-14th June 2018.
- 2018 Plenary speaker at International Society for Extracellular Vesicles ISEV18, and co-chair, Barcelona, 2-6th May 2018.
- 2018 Invited speaker at Bacterial Membrane Symposium, February 2018, Norwegian Arctic University Tromsø, Norway.
- 2015 Plenary speaker 9th European Zebrafish Meeting, Oslo. Use Of Zebrafish As Infection Model In Aquaculture.
- 2016 Invited lecturer PhD research school in Infection, Inflammation and Immunity, at Norwegian Arctic University Tromsø, Norway. Molecular and clinical aspects of infection, inflammation and immunity.

PUBLICATION LIST (peer review):

43. Thapa RK, **Winther-Larsen HC**, Diep DB, Tønnesen HH. 2020. Preformulation studies on novel garvicin KS peptides for topical applications. *European Journal of Pharmaceutical Sciences*. **151**:105333. <https://doi.org/10.1016/j.ejps.2020.105333>
42. Langlete P, Krabberød AK, **Winther-Larsen HC**. 2019. Vesicles from *Vibrio cholerae* contain AT-rich DNA and shorter mRNAs that do not correlate with their protein products. *Frontiers in Microbiol*. Accepted for publication.
41. Pöthig A, Ahmed S, **Winther-Larsen HC**, Guan S, Altmann PJ, Kudermann J, Andresen AMS, Gjøen T Åstrand OAH. 2018. Antimicrobial activity and cytotoxicity of Ag(I) and Au(I) pillarplexes. *Frontiers in chemistry*. Doi 10.3389/fchem.2018.00584.
40. Ulanova L, Pinheiro M, Vibe C, Nunes C, Misaghian D, Wilson S, Zhu K, Fenaroli F, **Winther-Larsen HC**, Reis S and Griffiths G. 2017. *Dis Aquat Organ*. 125:19.
39. Tandberg J, Lagos L, Ropstad E, Smistad G, Hiorth M and **Winther-Larsen HC**. 2018. The use of chitosan-coated membrane vesicles for immunization against salmonid rickettsial septicemia in an adult zebrafish model. *Zebrafish* 15:372.
38. Brenz Y, **Winther-Larsen HC**, Hagedorn M. 2018. Expanding Francisella models: Pairing up the soil amoeba Dictyostelium with aquatic Francisella. *International J Med Microbiol*. 308:32.
37. Kumar S, Tandberg J, Penesyan A, Elbourne L, Suarez Bosche N, Don E, Skadeberg E, Fenaroli F, Cole N, **Winther-Larsen HC**, Paulsen I. 2018. Dual transcriptomics of host-pathogen interaction of cystic fibrosis isolate *Pseudomonas aeruginosa* PASS1 with *Danio rerio*. 2018. *Frontiers Microbiology*. DOI: 10.3389/fcimb.2018.00406.
36. Karlsen C, Ottem KF, Brevik ØJ, Davey M, Sørum H and Winther-Larsen HC. 2017. The environmental and host-associated bacterial microbiota of Arctic seawater-farmed Atlantic salmon with ulcerative disorders. *J Fish Dis* **40**:1645.
35. Lagos L, Tandberg JI, Kashulin-Bekkelund A, Colquhoun DJ, Sørum H, **Winther-Larsen HC**. 2017. Isolation and characterization of serum extracellular vesicles (EVs) from Atlantic salmon infected with *Piscirickettsia salmonis*. *Proteomes*. 5:34. pii: E34. doi: 10.3390/proteomes5040034.
34. Lampe EO, Zingmark C, Tandberg JI, Thrane IM, Brudal E, Sjøstedt A and **Winther-Larsen HC**. 2017. *Francisella noatunensis* subspecies *noatunensis* *clpB* deletion mutant impairs development of francisellosis in a zebrafish model. *Vaccine*. 19:7264. IF 3.2.
33. Oliver C, Hernandez MA, Tandberg JI, Valenzuela KN, Lagos L, Haro R, Sánchez P, Ruiz PA, Sanhueza-Oyarzún C, Cortes MA, Lecumberri ATV, Artigues-Serra A, **Winther-Larsen HC**, Avendaño-Herrera R, Yañez AJ. 2017. The proteome of biologically active membrane vesicles from *Piscirickettsia salmonis* LF-89 type strain identifies plasmid-encoded putative toxins. *Frontiers in Cellular and Infection Microbiology*. 7:420. <https://doi.org/10.3389/fcimb.2017.00420>. IF 4.3.
32. Lagos L, Tandberg JI, Repnik U, Boysen P, Ropstad E, Varkay D, Paulsen IT and **Winther-Larsen HC**. 2017. *Clin Vaccine Immunol*. 24:e00557-16. Doi: 10.1128/CFI.00557-16.
31. Lagos L, Tandberg JT, Becker I, **Winther-Larsen HC**. 2017. Immunomodulatory properties of *Concholepas concholepas* hemocyanin against francisellosis in a zebrafish model. *Fish Shellfish Immunol*. pii: S1050-4648(17)30369-8. doi: 10.1016/j.fsi.2017.06.046. IF 3.3.

30. Tandberg JI, Oliver C, Lagos L, Gaarder M, Yáñez A, Ropstad E, **Winther-Larsen HC**. 2017. Membrane vesicles from *Piscirickettsia salmonis* induce protective immunity and reduce development of salmonid rickettsial septicemia in an adult zebrafish model. *Fish Shellfish Immunol.* 67:189-198. doi: 10.1016/j.fsi.2017.06.015. IF 3.3 (4 citations)
29. Brenz Y, Ohnezeit D, **Winther-Larsen HC**, Hagedorn M. 2017. Nramp1 and NrampB contribute to resistance against *Francisella* in *Dictyostelium*. *Frontiers in Cellular and Infection Microbiology*. <https://doi.org/10.3389/fcimb.2017.00282>.
28. Lampe EO, Brenz YP, Herrmann L, Repnik U, Griffiths GW, Zingmark C, Sjöstedt A, **Winther-Larsen HC** and Hagedorn M. 2015. Dissection of *Francisella* - host cell interactions in *Dictyostelium discoideum*. *Appl Environ Microbiol*.
27. Lampe EL, Tandberg JI, Rishovd A-L, **Winther-Larsen HC**. 2017. *Francisella noatunensis* ssp. *noatunensis igIC* deletion mutant protects adult zebrafish challenged with acute mortality dose of wild-type strain. *Dis Aquat Organ.* **123**:123.
26. Tandberg JI, Lagos LX, Langlete P, Berger E, Rishovd AL, Roos N, Varkay D, Paulsen IT, **Winther-Larsen HC**. 2016. Comparative analysis of membrane vesicles from three *Piscirickettsia salmonis* isolates reveals difference in vesicle characterisation. *PLOS One* 11: e0165099. Doi:10.1371/journal.pone.0165099.
25. Brudal E, Lampe EO, Reubsæet L, Roos N, Hegna IK, Thrane IM, Koppang EO & **Winther-Larsen HC**. 2015. Vaccination with outer membrane vesicles from *Francisella noatunensis* reduces development of francisellosis in a zebrafish model. *Fish Shellfish Immunol.* **42**:50.
24. Karlsen C, Ellingsen AB, Wiik-Nielsen C, **Winther-Larsen HC**, Colquhoun DJ and Sørum H. 2014. Host specificity and clade dependent distribution of putative virulence genes in *Moritella viscosa*. *Microbial Pathogenesis.* **77**:53.
23. Brudal E, Ulanova LS, Lampe EO, Rishovd A-L, Griffiths G, and **Winther-Larsen HC**. 2014. Establishment of three *Francisella* infections in zebrafish embryos at different temperatures. *Infection & Immunity.* **82**:2180.
22. Vestvik N, Rønneseth A, Kalgraff CAK, **Winther-Larsen HC**, Wergeland H and Haugland GT. 2013. *Francisella noatunensis* subsp. *noatunensis* replicates within Atlantic cod (*Gadus morhua* L.) leucocytes and inhibit respiratory burst activity. *Fish Shellfish Immunol.* **35**:725.
21. Brudal E, **Winther-Larsen HC**, Colquhoun DJ, and Duodu S. 2013. Evaluation of reference genes for reverse transcription quantitative PCR analyses of fish-pathogenic *Francisella* strains exposed to different growth conditions. *BMC Research Notes*, **6**:76.
20. Ford CG, Kolappan S, Phan HTH, **Winther-Larsen HC**, Waldor M and Craig L. 2012. Crystal Structures of CTX ϕ pIII Unbound and in Complex with *Vibrio cholerae* TolA Reveal Novel Interaction Interfaces. *J Biol Chem*, **287**:36258-72.
19. Bjelland AM, Sørum H, Tegegne DA, **Winther-Larsen HC**, Willassen NP and Hansen H. 2012. LitR of *Vibrio (Aliivibrio) salmonicida* is a salinity-sensitive quorum sensing regulator of phenotypes involved in host interactions and virulence. *Infection & Immunity*, **80**:1681-9.

18. Bjelland AM, Johansen R, Brudal E, Hansen H, **Winther-Larsen HC** and Sørum H. 2012. *Vibrio salmonicida* pathogenesis analyzed by experimental challenge of Atlantic salmon (*Salmo salar*). *Microbial Pathogenesis*. **52**. 77-84.
17. Bakkemo KR, Mikkelsen H, Bordevik M, Torgersen J, **Winther-Larsen HC**, Vanberg C, Olsen R, Johansen LH, Seppola M. 2011. Intracellular localisation and innate immune responses following *Francisella noatunensis* infection of Atlantic cod (*Gadus morhua*) macrophages. *Fish Shellfish Immunol*. **31**. 993-1004.
16. Egge-Jacobsen WM, Salomonsson EN, Aas FE, Forslund A-L, **Winther-Larsen HC**, Macellaro A, Kuoppa K, Oyston PCF, Titball RW, Thomas RM, Forsberg Å, Prior JL and Koomey M. 2011. O-linked glycosylation of the PilA pilin protein of *Francisella tularensis*: identification of the endogenous protein-targeting oligosaccharyltransferase and characterization of the native oligosaccharide. *J Bacteriol*. **193**. 5487-97.
15. Heiniger JW, **Winther-Larsen HC**, Pickles R, Koomey M and Wolfgang MC. 2010. Infection of human mucosal tissue by *Pseudomonas aeruginosa* requires sequential and mutually dependent virulence factors and a novel pilus-associated adhesin. *Cellular microbiology*. **12**. 1158-73.
14. Børud B, Aas F-E, Vik Å, **Winther-Larsen HC**, Egge-Jacobsen W, and Koomey M. 2010. Genetic, structural and antigenic analyses of glycan diversity in the O-linked protein glycosylation systems of human *Neisseria* species. *J Bacteriol*. **192**. 2816-29.
13. Salomonsson E, Forsberg Å, Roos N, Holz C, Maier B, Koomey M and **Winther-Larsen HC**. 2009. Functional analyses of pilin-like proteins from *Francisella* species: complementation of Type IV pilus phenotypes in *Neisseria gonorrhoeae*. *Microbiology*, **155**, 2546-59.
12. Næssan CL, Egge-Jacobsen W, Heiniger R, Wolfgang M, Røhr Å, Aas FE, **Winther-Larsen HC** and Koomey M. 2008. Genetic and structural characterization of *Neisseria gonorrhoeae* PptA, a phosphoethanolamine transferase targeting Type IV pili. 2008. *J Bacteriol*, **190**, 387-400.
- 11. Winther-Larsen HC**, Wolfgang MC, van Putten JMP, Roos N, Aas FE, Egge-Jakobsen W, Maier B, and Koomey M. 2007. *Pseudomonas aeruginosa* Type IV pilus expression in *Neisseria gonorrhoea*: effects of pilin subunit composition and function and organelle dynamics. *J Bacteriol*, **189**, 6676-85, 2007.
10. Aas FE, **Winther-Larsen HC**, Wolfgang M, Frye F, Løvold C, van Putten JMP, Roos N and Koomey M. 2006. Substitutions in the N-terminal alpha helical spine of *Neisseria gonorrhoeae* pilin affect Type IV pilus assembly, dynamics and associated functions. *Mol Microbiol* **63**, 69-85.
9. Aas FE, Egge-Jacobsen WM, **Winther-Larsen HC**, Løvold C, Hitchen PG, Dell A, and Koomey M. 2006. *Neisseria gonorrhoeae* type IV pili undergo multisite, hierarchical modifications with phosphoethanolamine and phosphocholine requiring an enzyme structurally related to LPS phosphoethanolamine transferases. *J Biol Chem* **22**, 27712-23.
8. **Winther-Larsen HC**, Wolfgang M, Dunham S, van Putten JP, Dorward D, Løvold C, Aas FE, and Koomey M. 2005. A conserved set of pilin-like molecules controls type IV pilus dynamics and organelle-associated functions in *Neisseria gonorrhoeae*. *Mol Microbiol*. **56**(4):903-17.
- 7. Winther-Larsen, H. C.** and Koomey, M. 2002. Transcriptional, chemosensory and cell contact-dependent regulation of Type IV pilus expression. *Current Opinion of Microbiology*. **5** (2), 173-178.

6. **Winther-Larsen, H. C.**, Hegge, F. T. Wolfgang, M, van Putten, J. P. M. Hayes, S. F. and Koomey, M. 2001. *Neisseria gonorrhoeae* PilV, a type IV pilus-associated protein essential to human epithelial cells adherence. PNAS **98**, 15276-15281.
5. **Winther-Larsen, H. C.**, Valand, B., Blatny, J. M., Brautaset, T. and Valla, S. 2000. *Pm* promoter expression mutants and their use in broad-host-range RK2 plasmid vectors. Metabolic Engineering. **2**, 92-103.
4. **Winther-Larsen, H. C.**, Josefsen, K., Brautaset, T. and Valla, S. 2000. Parameters affecting gene expression from the *Pm* promoter in Gram-negative bacteria. Metabolic Engineering. **2**, 70-91.
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