



KJM 5250 and KJM 9250
¹H NMR spectra on the AVneo400 spectrometer.
Version 3.1
Topspin 4.3



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AVneo400 ¹H NMR Spectra. Topspin 4.3.

1.0 Introduction

A series of aw coded ¹H NMR parameter sets have been created on the NEO-400 spectrometer. NEO400 spectra can be processed using **Topspin 3.5/3.6 or TS4**.

FT and **PK** (or **APK**) processing does not apply a line broadening factor. **EF** or **EFP** processing applies a line broadening factor (**LB**).

Resolution enhancement uses negative **LB** values. Try **LB** = -1.0 to -2.5 Hz with **GB** = 0.33, and **GFP** processing. Remember to reset **LB** and **GB** to their normal values (0.1 to 0.3 and 0 respectively) after **GFP** processing.

1.1 Presaturation Experiments

Continuous wave or excitation sculptured (ES) presaturation can be used to suppress ¹H NMR signals. The simplest of these techniques is continuous wave presaturation.

CW presaturation (PR) power levels (db settings) can be increased or decreased by subtracting or adding 3-12 db respectively. 6 db = a factor of 2. Bruker sometimes uses the **NOESYPR1D** pulse program to acquire presaturated ¹H NMR spectra and **QNMR** spectra.

The **ES** shaped pulse's excitation window can be decreased (softened) by doubling its shaped pulse time from 2000 usec to 4000 usec and halving its power by adding 6 db to that read in using the **getprosol** command.

1.2 Homonuclear Decoupling

Homonuclear decoupling experiments can be performed using Bruker's **zghd** or **zghd.2** pulse programmes which incorporate homonuclear decoupling during FD acquisition, or during both FID acquisition and the interpulse delay period (**d1**), respectively. The application of a decoupling radio frequency slightly increases the frequency of nearby signals. This effect is known as the Bloch-Seigert effect.

2.0 ¹H NMR experiments

2.1 ¹H spectrum with a 30 or 90 degree pulse

2.2 ¹H spectrum with CW presaturation

2.3 ¹H spectrum with F1 + F2 CW presaturation

2.4 ¹H spectrum with ES peak suppression

2.5 ¹H NOESYPR1D spectrum

**2.6 ¹H spectrum with homonuclear decoupling during FID acquisition
or during D1 + FID acquisition**

2.1 ^1H NMR with a 30 or 90 degree pulse

Parameter sets: **awproton30b** or **awproton90** (+ **getprosol**)

Pulse programmes: **zg30** or **zg** respectively

TD = 32 K, **SI** = 32 K.

SW = 16 ppm, **O1P** = 6 ppm

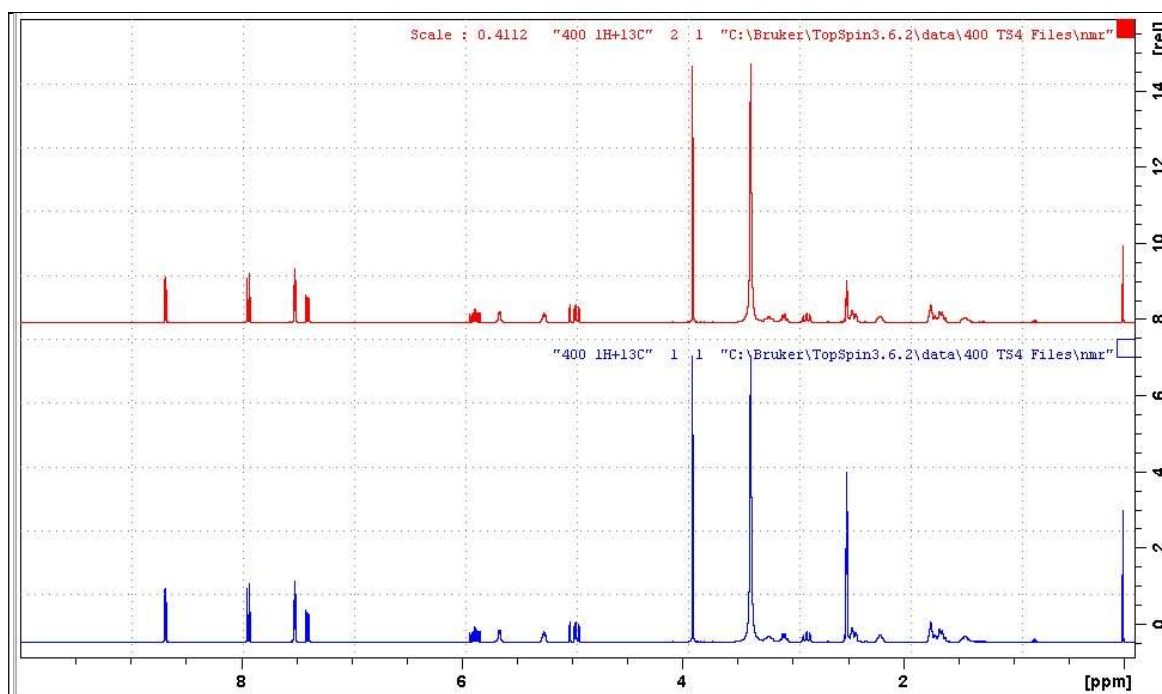
D1 = 2 - 5 sec or other time of your choice.

NS = any number, **DS** = 2, 4 or 8.

Type **ased** (enter) and review parameters used in the job.

Set **receiver gain** using **RGA** (*important!*)

Process with **FT** (no line broadening) or **EFP** (applies **LB**).



^1H NMR spectra of quinine in D_6 -DMSO using a 30 degree pulse (lower spectrum) or a 90 degree pulse (upper spectrum)..

2.2 ^1H NMR with CW presaturation

Parameter set: **awprotonpr (+ getprosol)**

Pulse programme: **zgpr**

TD = 32 K, **SI** = 32 K.

SW = 16 ppm, **O1P** = 6 ppm

O1 = frequency in Hz of the F1 signal to be presaturated
= spectral window mid-point. Check **SW** is wide enough.

PLW9(db) = F1 presaturation power applied during **D1**.

D1 = 2-5 sec or other time of your choice.

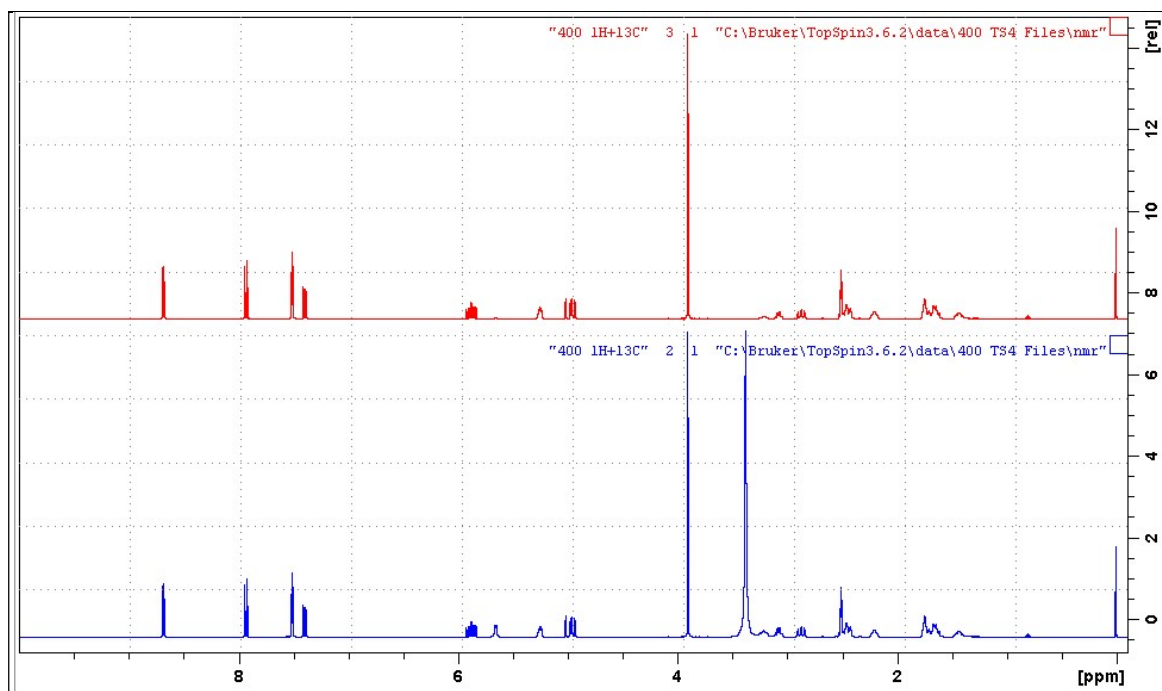
Type **ased** (enter) and review parameters used in the job.

Add (or subtract) 3-12 db to **PLW9** to decrease (or increase) the presaturation power.

6 db = a factor of 2. A larger attenuation setting decreases the power level.

Set **receiver gain** using **RGA** (*important!*).

Process with **EFP** (applies **LB**).



Lower: ^1H NMR spectrum of quinine in D_6 -DMSO.

Upper: ^1H NMR spectrum with CW presaturation of the HOD line at 3.37 ppm.

2.3 ^1H NMR with dual CW presaturation

Parameter set: **awprotonprf1prf2 (+ getprosol)**

Pulse programme: **awprotonprf1prf2**

TD = 32 K, **SI** = 32 K.

SW = 16 ppm, **O1P** = 6 ppm

O1 = frequency in Hz of the F1 signal to be presaturated
= spectral window mid-point. Check **SW** is wide enough.

O2 = frequency in Hz of the F2 signal to be presaturated.

PL9 = F1 presaturation power applied during **D1**.

PL21 = F2 presaturation power applied during **D1**.

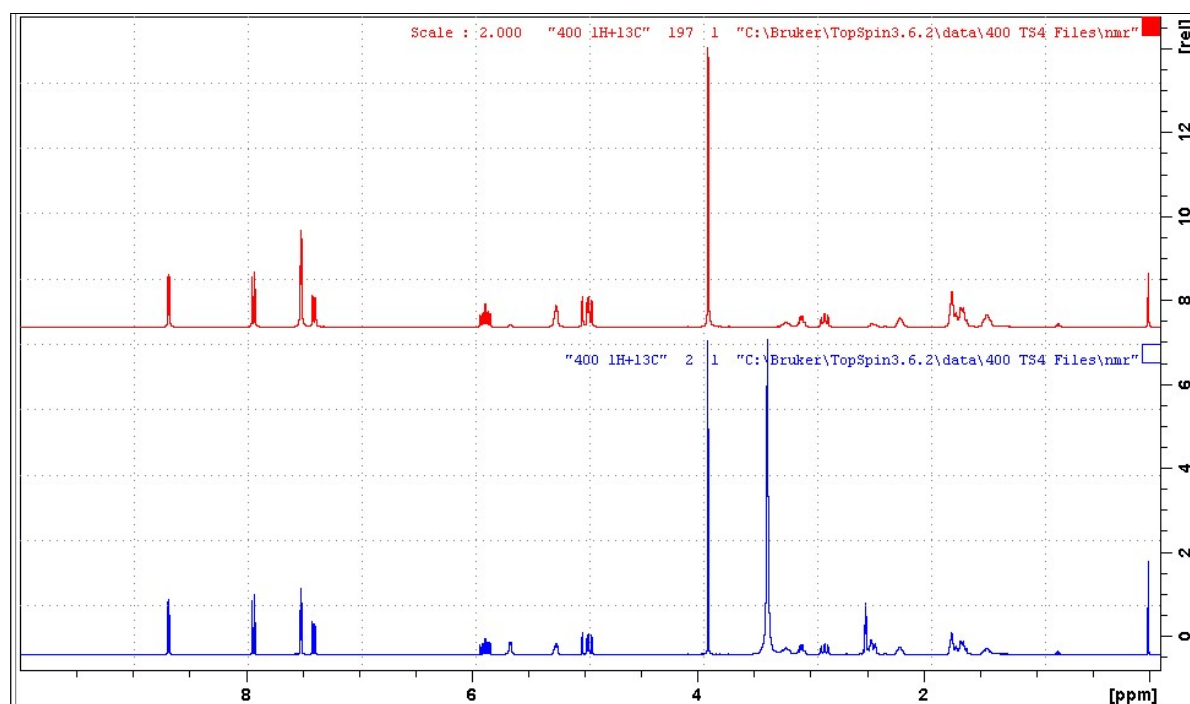
D1 = 2-5 sec or other time of your choice.

Type **ased** (enter) and review parameters used in the job.

Add (or subtract) 3-12 db to **PL9** and/or **PL21** to decrease (or increase) the presaturation power. 6 db = a factor of 2. A larger attenuation setting decreases the power level.

Set **receiver gain** using **RGA** (*important!*).

Process with **EFP** (applies **LB**).



Lower: ^1H NMR spectrum of quinine in D_6 -DMSO.

Upper: ^1H NMR spectrum with CW presaturation of the HOD (3.37 ppm) and DMSO (2.5 ppm) lines.

2.4 ^1H NMR with ES peak suppression

Parameter sets: **awprotones (+ getprosol)**

Pulse programmes: **zgesgp**

TD = 32 K, **SI** = 32 K.

SW = 16 ppm, **O1P** = 6 ppm

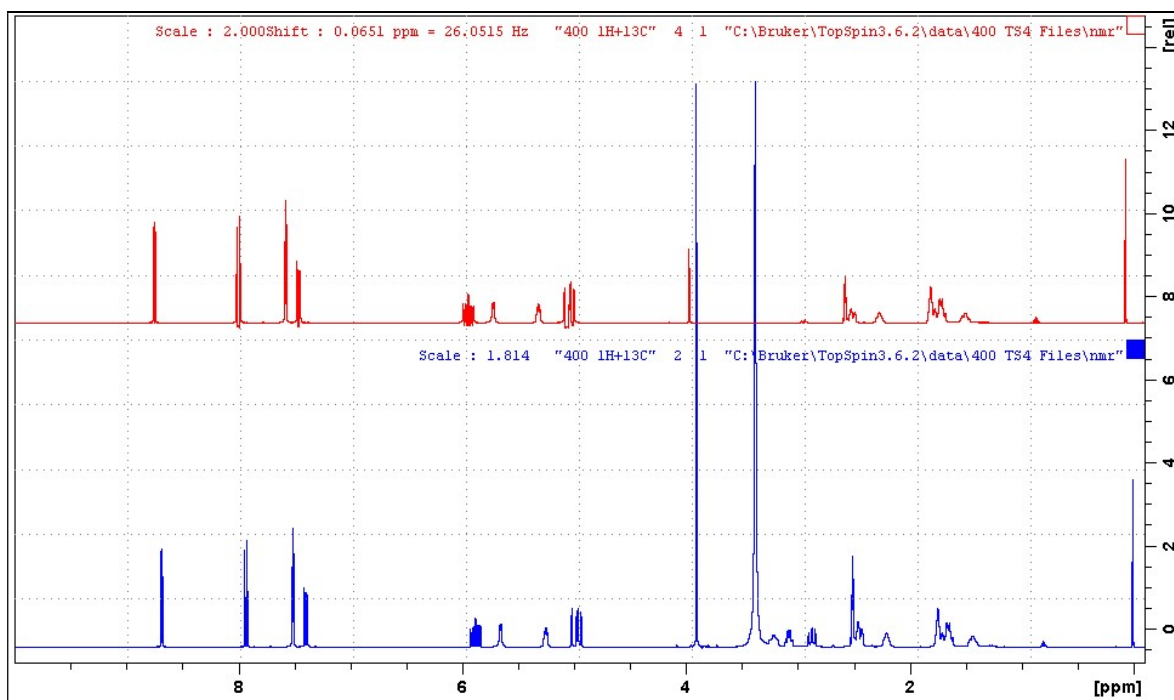
O1 = frequency in Hz of the F1 signal to be ES suppressed
= spectral window mid-point. Check **SW** is wide enough.

D1 = 2 sec or other time of your choice.

Type **ased** (enter) and review parameters used in the job. Verify gradients are OK.
Check **P12** = 2000 usec, **SPNAM1** = **squa100.1000**.

Set **receiver gain** using **RGA** (*important!*).

Process with **EFP** (applies **LB**).



Lower: ^1H NMR spectrum of quinine in $\text{D}_6\text{-DMSO}$.

Upper: ^1H NMR spectrum with ES suppression of the HOD line at 3.37 ppm. The ES spectrum is slightly offset relative to the ^1H NMR spectrum.

400 MHz ES suppression of the 3.37 ppm HOD line significantly or completely reduces the peak areas of quinine signals located 0.5-0.8 ppm either side of the HOD line.

2.5 NOESYPR1D Spectrum with CW Presaturation

Parameter set: **awnoesypr1d (+ getprosol)**

Pulse programme: **noesypr1d**

TD = 32 K, **SI** = 32 K.

SW = 16 ppm, **O1P** = 6 ppm

O1 = frequency in Hz of the F1 signal to be presaturated

= spectral window mid-point. Check **SW** is wide enough.

PLW9 = prosol Table linked presaturation power applied during **D1**.

D8 = 0.05 sec or other time of your choice

D1 = 2-5 sec or other time of your choice.

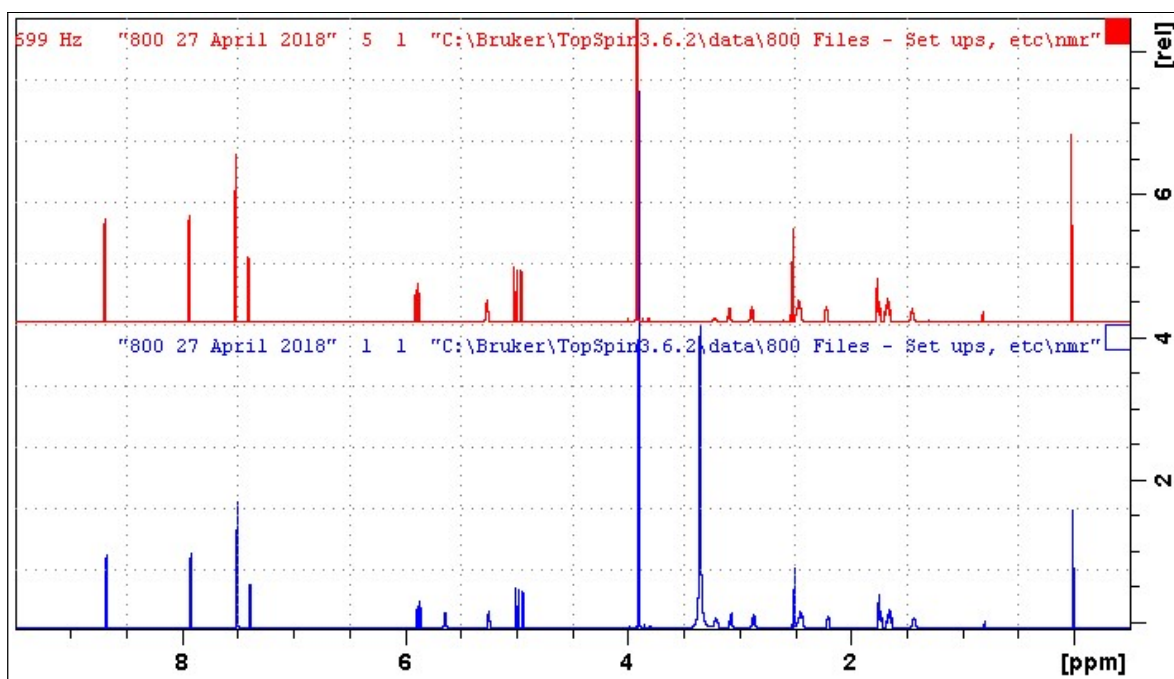
Type **ased** (enter) and review parameters used in the job.

Add (or subtract) 3-12 db to **PLW9(db)** to decrease (or increase) the presaturation power.

6 db = a factor of 2. A larger attenuation setting decreases the power level.

Set **receiver gain** using **RGA** (*important!*).

Process with **EFP** (applies **LB**).



Lower: ^1H NMR spectrum of quinine in D_6 -DMSO.

Upper: NOESYPR1D spectrum with CW presaturation of the HOD line at 3.37 ppm

2.6 ^1H spectrum with homonuclear decoupling during FID acquisition or during D1 + FID acquisition

Parameter set: **awprotonhd** or **awprotonhd.2** (+ **getprosol**)

Pulse programme: **zghd** or **zghd.2**

Prior to setting up a **homonuclear decoupling** experiment determine the frequency in **Hz** of the signal to be decoupled in a standard ^1H NMR spectrum.

Type **O2** (enter) and enter the frequency in Hz of the signal to be decoupled.

Type **eda** (enter) and adjust acquisition parameters as required.

SW = 16 ppm, **O1P** = 6 ppm, **TD** = 32 K points or other values of your choice.

NS = multiple of 2, 4 or 8, **DS** = 2, 4 or 8.

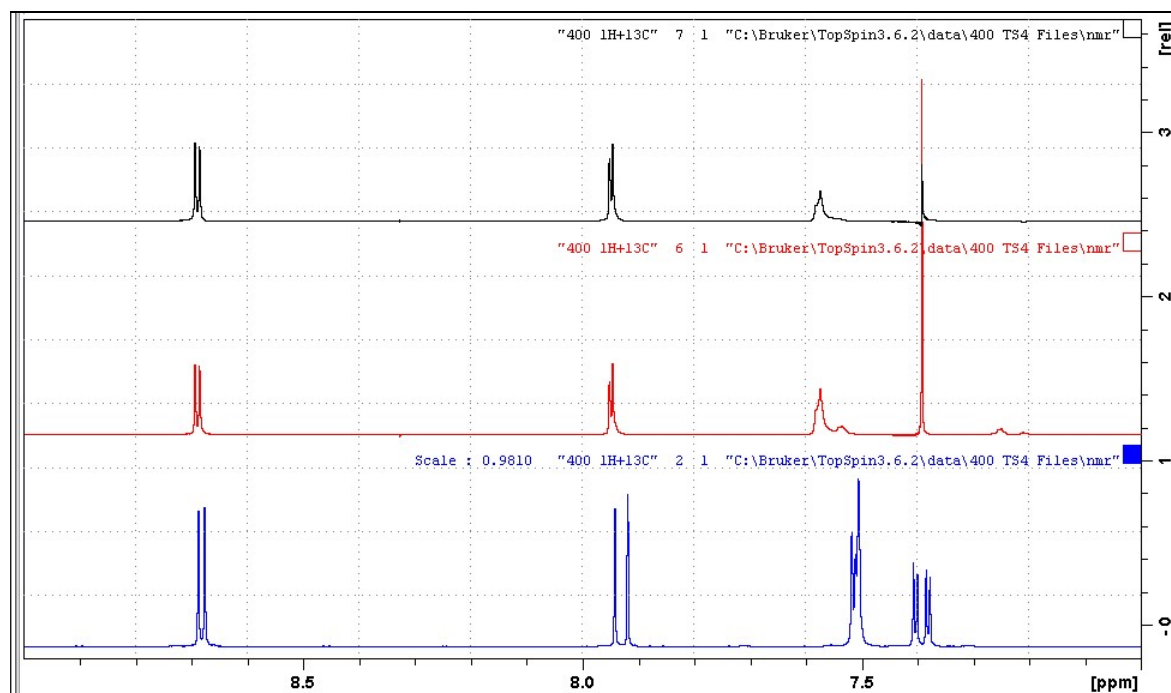
D1 = repetition delay = 2-3 sec or other time of your choice.

Type **ased** (enter) and review other parameters including the **PL24** power level.

Do not adjust the **PL1** or **PL2** power levels.

Set **receiver gain** using **RGA** (*important!*).

Process with **EF** or **EFP** (applies **LB** = 0.1, 0.3 Hz or other values of your choice)
or **FT** (no line broadening factor applied).



Lower: The 7-9 ppm region of the ^1H NMR spectrum of quinine in $\text{D}_6\text{-DMSO}$.

Center: 7-9 ppm region with homonuclear decoupling at 7.39 ppm during FID acquisition.

Upper: 7-9 ppm region with homonuclear decoupling at 7.39 ppm during D1 + FID acquisition.